Boundary-Specific Remodeling Defines Divergent TLS Maturation in Colorectal Cancer

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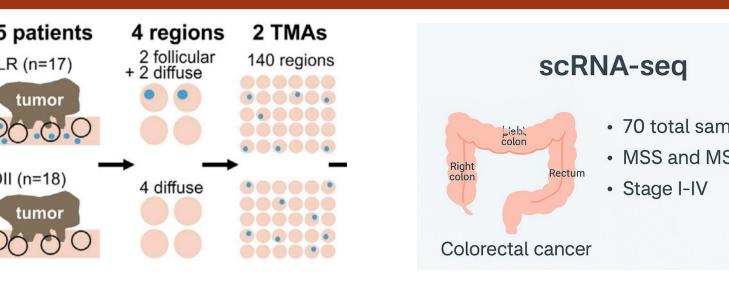
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Introduction and Aim

- •Immunotherapy has shown major success in cancers such as melanoma and lung cancer, but most colorectal cancers (CRC) do not respond effectively.
- •Tertiary lymphoid structures (TLSs) are organized clusters of immune cells that form within tumors and can enhance local antitumor immunity.
- •The maturation process and microenvironmental organization of TLSs in colorectal cancer remain poorly understood.
- •Hypothesis: TLS maturation involves coordinated structural and gene expression remodeling, which differs between immunotherapy-responsive, colorectal-like responsive (CLR), and non-responsive, diffuse immune-inactive (DII), colorectal cancers.

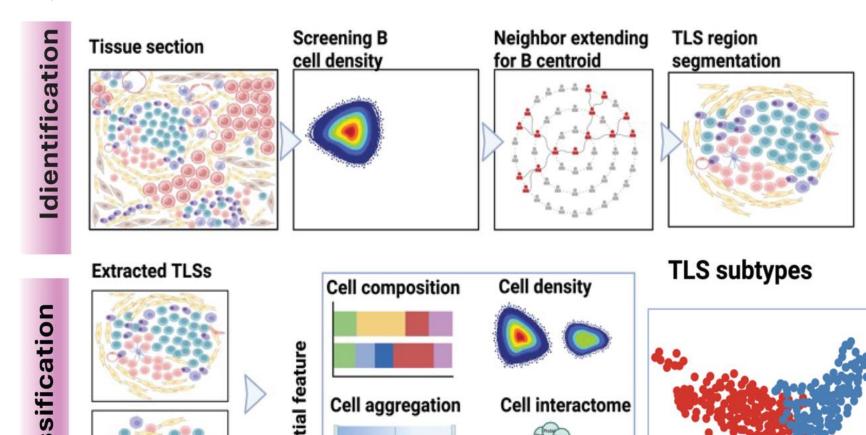
Data & Methods

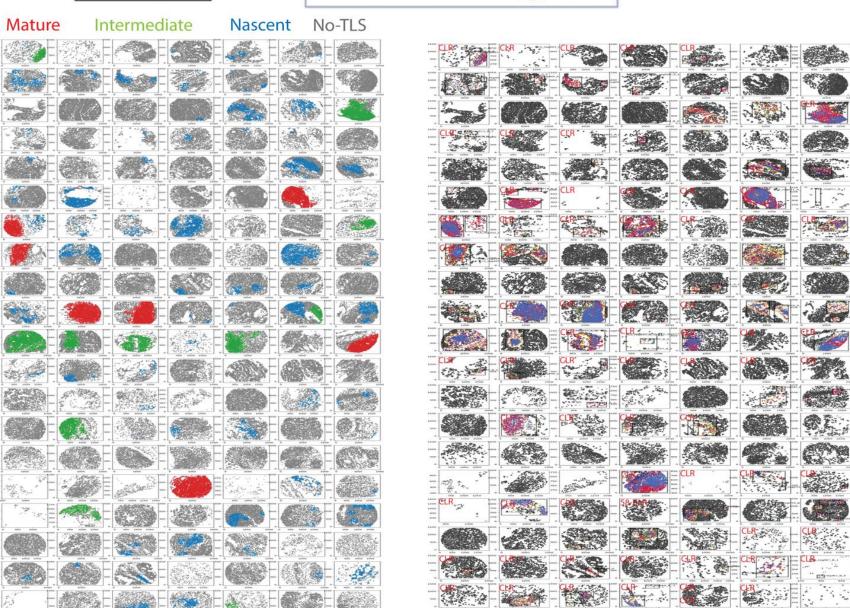


70 total samples MSS and MSI tumors

Figure 1. Multi-omics datasets for TLS analysis in colorectal

Spatial proteomics data and single-cell RNA sequencing data were integrated to map structural and transcriptional features of tertiary lymphoid structure maturation.





TLS analysis in colorectal cancer. Spatial proteomics data and single-cell RNA sequencing data were integrated to map structural and transcriptional features of tertiary lymphoid structure

Figure 3. Multi-omics datasets for

Spatial proteomics data and single-cell

RNA sequencing data were integrated to

map structural and transcriptional

features of tertiary lymphoid structure

MaxFuse

scRNA-seq

Figure 4. Multi-omics datasets for

Fused output

maturation.

maturation.

TLS analysis in colorectal cancer.

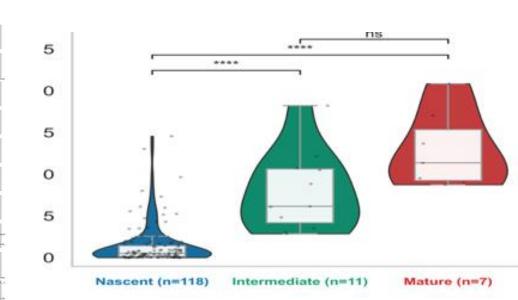


Figure 5: TLS number and area

Quantification of TLS number and area per maturation stage demonstrates expansion progressive consolidation of TLS structures.

Figure 2. Identification and classification of TLS maturation states.

Workflow for TLS detection and classification. TLS regions were identified by screening B-cell density, expanding neighbors around B-cell centroids, and segmenting contiguous clusters into TLS regions. Extracted TLSs were further characterized by cell density, aggregation, composition, and interactome features. Representative TLSs across TMA cohorts A and B illustrate classification into nascent (blue), intermediate (green), and mature (red) maturation states. Cell composition derived by the classification algorithm provides reference distributions of immune subsets across TLS subtypes.

Results

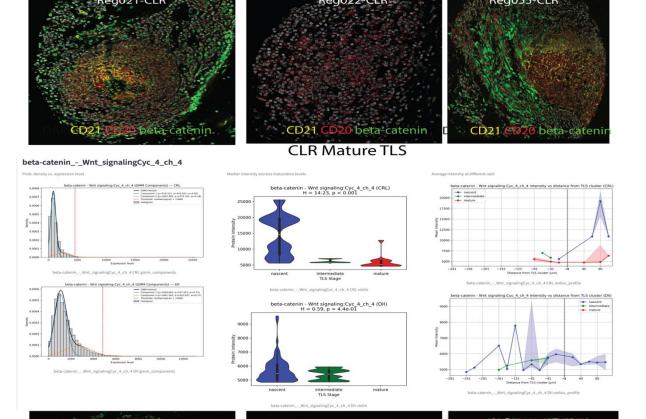
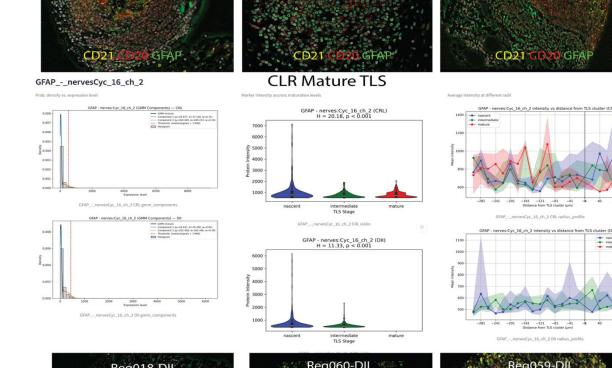


Figure 6. β -catenin expression.

CLR TLSs show **\(\beta\)-catenin suppression inside** TLSs and enrichment outside boundaries. indicating boundary-specific Wnt signaling, whereas DII TLSs lack clear spatial gradients and display diffuse β-catenin expression.



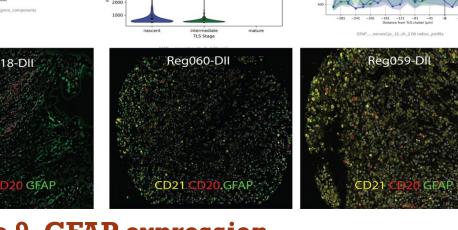


Figure 9. GFAP expression. CLR TLSs show strong GFAP enrichment inside mature TLSs, indicating localized stromal activation, whereas DII TLSs display diffuse, unpatterned GFAP expression lacking spatial organization.

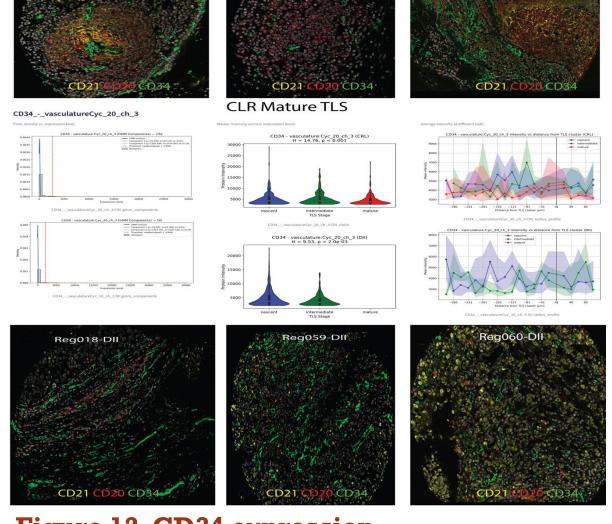
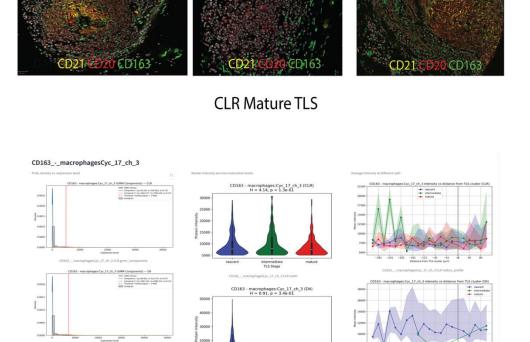


Figure 12. CD34 expression.

CLR TLSs show reduced CD34+ vasculature inside mature TLSs, indicating vascular exclusion, whereas DII TLSs display higher, diffuse CD34 expression consistent with persistent vascular activity.



Figure 7. Collagen IV expression. Mature CLR TLSs show structured Collagen IV deposition at TLS boundaries, indicating extracellular matrix remodeling, while DII TLSs display diffuse, unorganized Collagen IV expression lacking boundary formation.



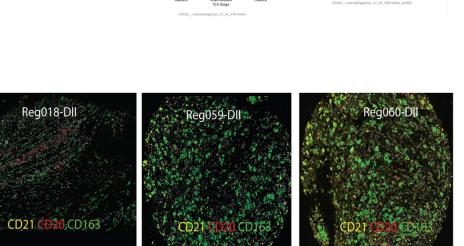


Figure 10. CD163 expression. CD163⁺ macrophages are enriched in

nascent DII TLSs and decline with maturation, while CLR TLSs show consistently low, diffuse expression across stages.

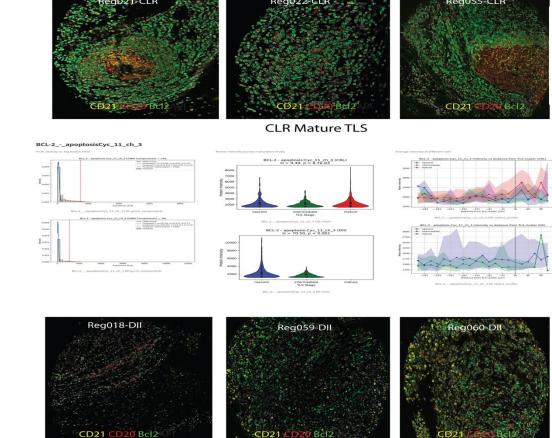


Figure 13. BCL-2 expression. Mature CLR TLSs show a BCL-2+ survival rim outside TLS boundaries, indicating compartmentalized apoptotic regulation, while DII TLSs lack this spatial organization.

Figure 8. α SMA expression Mature CLR TLSs show strong αSMA+ stromal encapsulation around TLS boundaries. whereas DII TLSs lack structured aSMA organization and display diffuse, low expression.

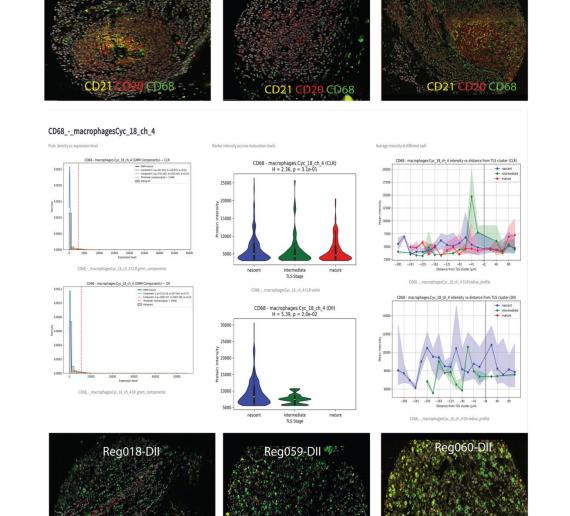


Figure 11. CD68 expression. CD68⁺ macrophages are enriched in nascent DII TLSs and decline with maturation, while CLR TLSs maintain uniformly low expression without boundary-specific organization.

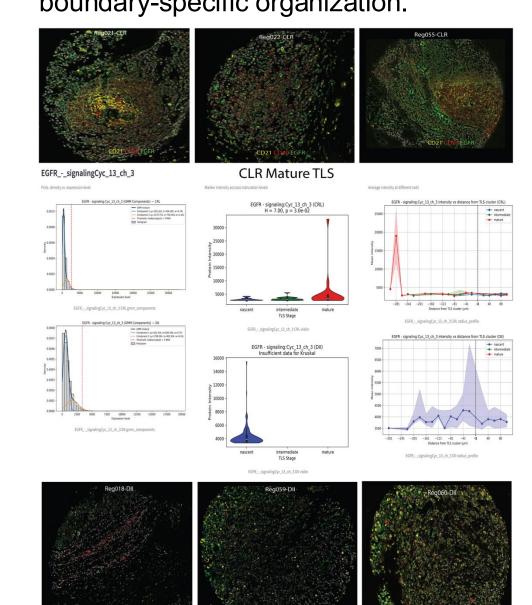


Figure 14. EGFR expression. Mature CLR TLSs show a sharp **EGFR** peak at the TLS boundary, indicating boundary-restricted signaling, whereas DII TLSs display diffuse, low EGFR expression without spatial organization.

Results (Cont.)

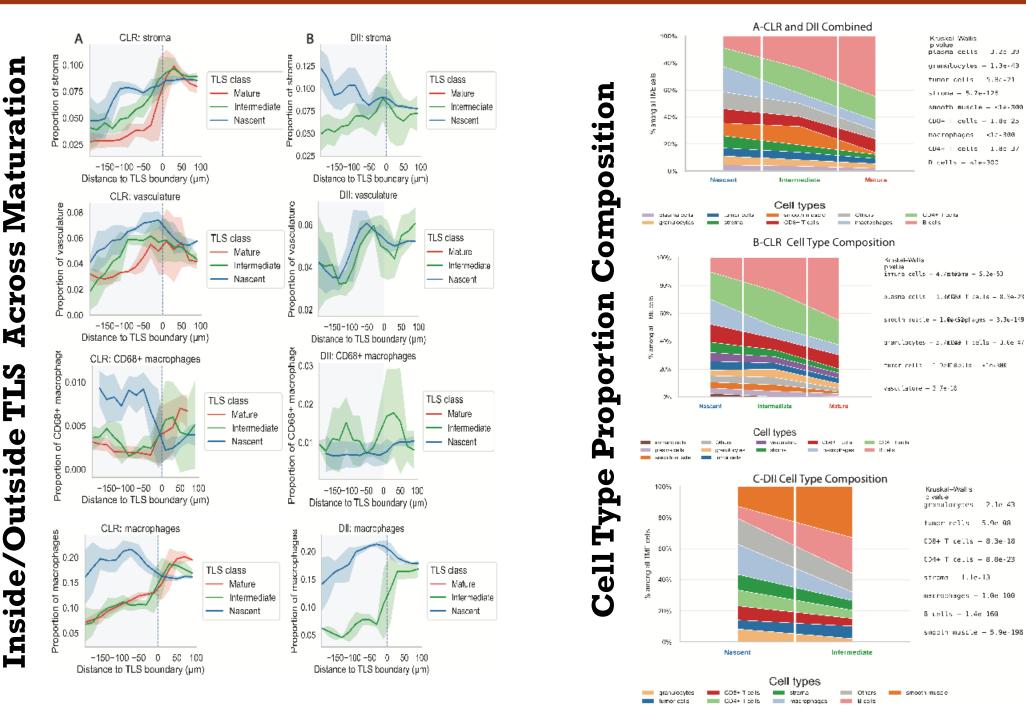
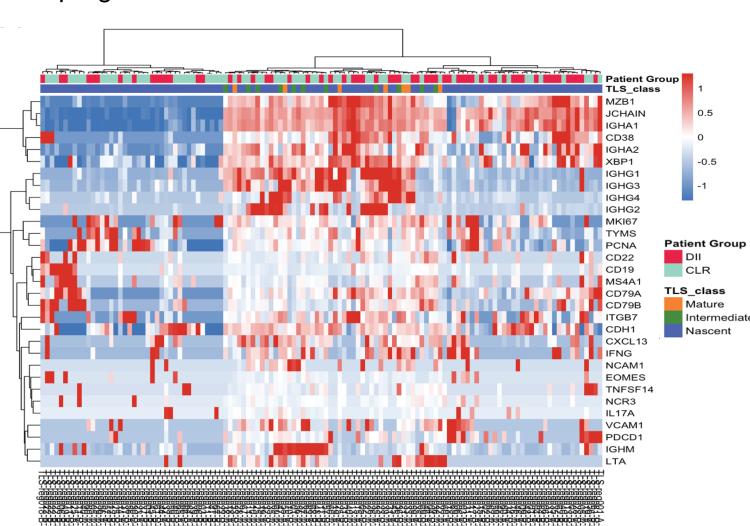


Figure 15. Cellular composition and spatial remodeling across TLS maturation. Gradient plots (left) show spatial distributions of stroma, vasculature, and macrophages relative to the TLS boundary in **CLR** and **DII** patients, with lines indicating TLS stages nascent (blue), intermediate (green), and mature (red). CLR TLSs show organized boundary enrichment of stromal and macrophage populations, whereas DII TLSs remain diffuse. Stacked barplots (right) display overall cell-type composition changes across maturation, revealing decreased B and CD4⁺ T cells and increased plasma cells, macrophages, and stroma with TLS progression.

Figure 16. Gene expression patterns across TLS maturation in CRC

showing key genes across TLSs from CLR (teal) and DII (red) CLR TLSs display strong induction of differentiation B-cell (MZB1 IGHA/IGHG) and immune activation (CXCL13, IFNG, PDCD1) with maturation, whereas DII TLSs show unstructured transcriptional programs.



Conclusions & Discussion

Conclusion

- TLS maturation in colorectal cancer involves coordinated stromal, vascular, and immune remodeling.
- Mature CLR TLSs form structured immune niches with αSMA+/Collagen IV+ capsules, boundary-restricted signaling (βcatenin, EGFR, BCL-2), and vascular exclusion.
- **DII TLSs** remain diffuse, lacking spatial and transcriptional compartmentalization.
- MaxFuse Integrated spatial proteomics and scRNA-seq revealed a shift from proliferative (MKI67⁺) nascent TLSs to plasma cell-rich (MZB1⁺, XBP1⁺) mature TLSs with enhanced immune effector activity.

Discussion

- Further work is needed to validate marker—cell type specificity and replicate findings in independent datasets.
- Future studies should integrate matched genomic and proteomic data from the same patients for deeper mechanistic insight.

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References

- Schürch, C.M., Bhate, S.S., Barlow, G.L., et al. Coordinated cellular neighborhoods orchestrate antitumoral immunity at the colorectal cancer invasive front. Cell 182, 1341–1359.e19 (2020).
- Pelka, K., Hofree, M., Chen, J.H., et al. Spatially organized multicellular immune hubs in human colorectal cancer. Cell 184, 4734–4752.e20 (2021)
- Gao, L., Bhaduri, A., et al. MaxFuse: Cross-modal integration of spatial and single-cell omics via graph matching and manifold alignment. Nature Methods 20, 1183–1193 (2023). • Sautès-Fridman, C., Petitprez, F., Calderaro, J., & Fridman, W.H. TLS in the era of cancer immunotherapy. Nature Reviews Cancer 19, 307–325 (2019).
- Cabrita, R., Lauss, M., Sanna, A., et al. Tertiary lymphoid structures improve immunotherapy and survival in melanoma. Nature 577, 561–565 (2020). • Helmink, B.A., Reddy, S.M., Gao, J., et al. B cells and tertiary lymphoid structures promote immunotherapy response. Nature 577, 549–555 (2020). • Bruno, T.C. New roles for TLS in antitumor immunity and response to immunotherapy. Frontiers in Immunology 11, 611928 (2021).